

Package: survinger (via r-universe)

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Title Design-Adjusted Inference for Pathogen Lineage Surveillance

Version 0.1.1

Description Provides tools for optimizing sequencing resource allocation and estimating pathogen lineage prevalence under real-world genomic surveillance conditions. Implements constrained allocation optimization for limited sequencing capacity across multiple regions and sample sources. Includes Horvitz-Thompson and post-stratified estimators that account for unequal sequencing rates, delay-adjusted nowcasting for right-censored reporting data, and combined design-weighted delay-corrected inference with uncertainty propagation.

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URL <https://github.com/CuiweiG/survinger>,
<https://cuiweig.github.io/survinger>

BugReports <https://github.com/CuiweiG/survinger/issues>

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`glance.surv`*One-row summary of survinger model*

Description

One-row summary of survinger model

Usage

```
## S3 method for class 'surv_prevalence'  
glance(x, ...)  
  
## S3 method for class 'surv_delay_fit'  
glance(x, ...)  
  
## S3 method for class 'surv_adjusted'  
glance(x, ...)
```

Arguments

`x` A survinger result object.
`...` Additional arguments (currently unused).

Value

A single-row tibble with key summary statistics.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)  
d <- surv_design(sim$sequences, ~ region,  
                  sim$population[c("region", "seq_rate")], sim$population)  
prev <- surv_lineage_prevalence(d, "BA.2.86")  
glance(prev)
```

`plot.surv`*Plot methods for survinger objects*

Description

Visualize surveillance design, allocation, prevalence estimates, delay distributions, nowcasts, and adjusted estimates.

Usage

```
## S3 method for class 'surv_design'
plot(x, ...)

## S3 method for class 'surv_allocation'
plot(x, ...)

## S3 method for class 'surv_prevalence'
plot(x, ...)

## S3 method for class 'surv_delay_fit'
plot(x, ...)

## S3 method for class 'surv_nowcast'
plot(x, ...)

## S3 method for class 'surv_adjusted'
plot(x, ...)
```

Arguments

x	A survinger object.
...	Additional arguments (currently unused).

Value

A ggplot2 object.

print.surv_adjusted *Combined design-weighted and delay-adjusted prevalence*

Description

Simultaneously corrects for unequal sequencing rates and right-truncation from reporting delays.

Usage

```
## S3 method for class 'surv_adjusted'
print(x, ...)

## S3 method for class 'surv_adjusted'
as.data.frame(x, ...)

surv_adjusted_prevalence(
  design,
  delay_fit,
```

```

    lineage,
    time = "epiweek",
    prevalence_method = "hajek",
    nowcast_method = "direct",
    conf_level = 0.95,
    bootstrap_n = 0L
  )

```

Arguments

x	Object to print.
...	Additional arguments (unused).
design	A surv_design object.
delay_fit	A surv_delay_fit object.
lineage	Character. Target lineage.
time	Character. Default "epiweek".
prevalence_method	Character. Default "hajek".
nowcast_method	Character. Default "direct".
conf_level	Numeric. Default 0.95.
bootstrap_n	Integer. 0 for delta method, >0 for bootstrap. Default 0.

Value

Invisibly returns the input object.

A surv_adjusted object.

Examples

```

sim <- surv_simulate(n_regions = 3, n_weeks = 12, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                sim$population[c("region", "seq_rate")], sim$population)
delay <- surv_estimate_delay(d)
adj <- surv_adjusted_prevalence(d, delay, "BA.2.86")
print(adj)

```

print.surv_allocation *Optimize sequencing allocation across strata*

Description

Given fixed total sequencing capacity, finds the optimal allocation across strata that minimizes a specified objective function.

Usage

```
## S3 method for class 'surv_allocation'
print(x, ...)

## S3 method for class 'surv_allocation'
as.data.frame(x, ...)

surv_optimize_allocation(
  design,
  objective = c("min_mse", "max_detection", "min_imbalance"),
  total_capacity,
  budget = NULL,
  min_per_stratum = 2L,
  target_lineage = NULL,
  target_prevalence = 0.01,
  cost_col = NULL
)
```

Arguments

x	Object to print.
...	Additional arguments (unused).
design	A surv_design object.
objective	Character. One of "min_mse", "max_detection", or "min_imbalance".
total_capacity	Integer. Total sequences available.
budget	Numeric or NULL. Optional budget constraint.
min_per_stratum	Integer. Minimum per stratum. Default 2.
target_lineage	Character. Required for "max_detection".
target_prevalence	Numeric. Assumed prevalence for detection. Default 0.01.
cost_col	Character or NULL. Column name for per-sequence cost.

Value

Invisibly returns the input object.
 A surv_allocation object.

Examples

```
sim <- surv_simulate(n_regions = 4, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
  sim$population[c("region", "seq_rate")], sim$population)
a <- surv_optimize_allocation(d, "min_mse", total_capacity = 500)
print(a)
```

print.surv_delay_fit *Estimate reporting delay distribution*

Description

Fits a parametric distribution to the delay between sample collection and sequence reporting, accounting for right-truncation.

Usage

```
## S3 method for class 'surv_delay_fit'
print(x, ...)

surv_estimate_delay(
  design,
  distribution = c("negbin", "poisson", "lognormal", "nonparametric"),
  strata = NULL,
  max_delay = 60L,
  ref_date = NULL
)
```

Arguments

x	Object to print.
...	Additional arguments (unused).
design	A surv_design object with both collection and report dates.
distribution	Character: "negbin" (default), "poisson", "lognormal", or "nonparametric".
strata	Optional one-sided formula for delay stratification.
max_delay	Integer. Maximum plausible delay in days. Default 60.
ref_date	Date. Reference date for right-truncation. Default is max report date in data.

Value

Invisibly returns the input object.

A surv_delay_fit object.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 12, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
fit <- surv_estimate_delay(d)
print(fit)
```

```
print.surv_design      Create a genomic surveillance design object
```

Description

Constructs a survey design object tailored for pathogen genomic surveillance, encoding stratification structure, sequencing rates, and population information needed for design-weighted inference.

Usage

```
## S3 method for class 'surv_design'
print(x, ...)

## S3 method for class 'surv_design'
summary(object, ...)

## S3 method for class 'summary.surv_design'
print(x, ...)

surv_design(
  data,
  strata,
  sequencing_rate,
  population,
  date_collected = "collection_date",
  date_reported = "report_date",
  lineage = "lineage",
  source_type = NULL,
  source_config = NULL
)
```

Arguments

x	Object to print or summarize.
...	Additional arguments (unused).
object	A surv_design object to summarize.
data	Data frame of individual sequence records.
strata	One-sided formula specifying stratification variables (e.g., ~ region or ~ region + source_type).
sequencing_rate	Either a one-sided formula (~ n_seqenced / n_positive), a named numeric vector, or a data frame with strata columns and seq_rate.
population	Data frame with one row per stratum, containing stratification variables and population-level denominators.
date_collected	Column name for collection date. Default "collection_date".

date_reported Column name for report date. Default "report_date". Set NULL if unavailable.
 lineage Column name for lineage. Default "lineage".
 source_type Column name for sample source. Default NULL.
 source_config Optional tibble of per-source characteristics.

Value

Invisibly returns the input object.
 A summary list of class `summary.surv_design`.
 Invisibly returns the input object.
 An object of class `surv_design`.

See Also

[surv_simulate\(\)](#), [surv_lineage_prevalence\(\)](#), [surv_optimize_allocation\(\)](#)

Examples

```

sim <- surv_simulate(n_regions = 3, n_weeks = 8, seed = 42)
design <- surv_design(
  data = sim$sequences,
  strata = ~ region,
  sequencing_rate = sim$population[c("region", "seq_rate")],
  population = sim$population
)
print(design)

```

print.surv_nowcast *Nowcast lineage counts correcting for reporting delays*

Description

Nowcast lineage counts correcting for reporting delays

Usage

```

## S3 method for class 'surv_nowcast'
print(x, ...)

## S3 method for class 'surv_nowcast'
as.data.frame(x, ...)

surv_nowcast_lineage(
  design,
  delay_fit,

```

```

lineage = NULL,
time = "epiweek",
horizon = 4L,
ref_date = NULL,
method = c("direct", "em")
)

```

Arguments

x	Object to print.
...	Additional arguments (unused).
design	A surv_design object.
delay_fit	A surv_delay_fit object.
lineage	Character or NULL. Target lineage.
time	Character. Default "epiweek".
horizon	Integer. Recent periods to nowcast. Default 4.
ref_date	Date or NULL. Default max report date.
method	Character: "direct" (default) or "em".

Value

Invisibly returns the input object.

A surv_nowcast object.

Examples

```

sim <- surv_simulate(n_regions = 3, n_weeks = 12, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
fit <- surv_estimate_delay(d)
nc <- surv_nowcast_lineage(d, fit, "BA.2.86")
print(nc)

```

print.surv_prevalence *Estimate lineage prevalence with design weights*

Description

Estimates the prevalence of a specified pathogen lineage over time, correcting for unequal sequencing rates across strata.

Usage

```
## S3 method for class 'surv_prevalence'  
print(x, ...)  
  
## S3 method for class 'surv_prevalence'  
as.data.frame(x, ...)  
  
surv_lineage_prevalence(  
  design,  
  lineage,  
  time = "epiweek",  
  method = c("hajek", "horvitz_thompson", "poststratified"),  
  conf_level = 0.95,  
  min_obs = 5L  
)
```

Arguments

<code>x</code>	Object to convert.
<code>...</code>	Additional arguments (unused).
<code>design</code>	A <code>surv_design</code> object.
<code>lineage</code>	Character. Target lineage name.
<code>time</code>	Character. Time aggregation: "epiweek", "month", "date", or a column name. Default "epiweek".
<code>method</code>	Character. Estimation method: "hajek" (default), "horvitz_thompson", or "poststratified".
<code>conf_level</code>	Numeric. Confidence level. Default 0.95.
<code>min_obs</code>	Integer. Minimum observations per time period. Default 5.

Value

Invisibly returns the input object.

A `data.frame`.

A `surv_prevalence` object.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)  
d <- surv_design(sim$sequences, ~ region,  
                 sim$population[c("region", "seq_rate")], sim$population)  
prev <- surv_lineage_prevalence(d, "BA.2.86")  
print(prev)
```

sarscov2_surveillance *Example SARS-CoV-2 genomic surveillance data*

Description

Simulated genomic surveillance dataset with 5 regions, 26 weeks, highly unequal sequencing rates (15% to 0.5%), three sample sources, and negative binomial reporting delays. Contains known ground truth for benchmarking.

Usage

```
sarscov2_surveillance
```

Format

A named list with four elements:

sequences Tibble of sequence records: sequence_id, region, source_type, lineage, collection_date, report_date, epiweek.

population Tibble with one row per region: region, n_positive, n_sequenced, seq_rate, pop_total.

truth Tibble of true lineage prevalence by region and week.

parameters List of simulation parameters.

Source

Simulated using `surv_simulate(seed = 20240101)`.

Examples

```
data(sarscov2_surveillance)
head(sarscov2_surveillance$sequences)
sarscov2_surveillance$population
```

surv_bind

Combine multiple prevalence estimates

Description

Bind results from different lineages or methods into a single tibble for comparison plots and tables.

Usage

```
surv_bind(...)
```

Arguments

... surv_prevalence objects to combine.

Value

A tibble with a source column identifying each input.

See Also

[surv_lineage_prevalence\(\)](#), [surv_sensitivity\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
p1 <- surv_lineage_prevalence(d, "BA.5")
p2 <- surv_lineage_prevalence(d, "XBB.1.5")
combined <- surv_bind(p1, p2)
head(combined)
```

surv_compare_allocations

Compare multiple allocation strategies

Description

Compare multiple allocation strategies

Usage

```
surv_compare_allocations(
  design,
  strategies = c("equal", "proportional", "min_mse", "max_detection", "min_imbalance"),
  total_capacity,
  target_prevalence = 0.01,
  ...
)
```

Arguments

design A surv_design object.

strategies Character vector. Default includes all built-in.

total_capacity Integer. Total sequences.

target_prevalence Numeric. For detection objective.

... Passed to [surv_optimize_allocation\(\)](#).

Value

A tibble comparing strategies.

Examples

```
sim <- surv_simulate(n_regions = 4, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_compare_allocations(d, total_capacity = 200)
```

surv_compare_estimates

Compare weighted vs naive prevalence estimates

Description

Side-by-side plot showing the impact of design correction.

Usage

```
surv_compare_estimates(weighted, naive, title = NULL)
```

Arguments

weighted	A surv_prevalence object (design-weighted).
naive	A surv_prevalence object (unweighted).
title	Character or NULL. Plot title.

Value

A ggplot2 object.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
w <- surv_lineage_prevalence(d, "BA.2.86")
n <- surv_naive_prevalence(d, "BA.2.86")
surv_compare_estimates(w, n)
```

surv_design_effect *Compute design effect over time*

Description

Compute design effect over time

Usage

```
surv_design_effect(weighted, naive)
```

Arguments

weighted A surv_prevalence object.
naive A surv_prevalence object.

Value

A tibble with time, deff, and bias_correction columns.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
w <- surv_lineage_prevalence(d, "BA.2.86")
n <- surv_naive_prevalence(d, "BA.2.86")
surv_design_effect(w, n)
```

surv_detection_probability

Variant detection probability under current design

Description

Variant detection probability under current design

Usage

```
surv_detection_probability(
  design,
  true_prevalence,
  delay_fit = NULL,
  n_periods = 1L,
  detection_threshold = 1L
)
```

Arguments

design A surv_design object.
true_prevalence Numeric in (0,1).
delay_fit Optional surv_delay_fit.
n_periods Integer. Accumulation periods. Default 1.
detection_threshold Integer. Min detections. Default 1.

Value

A list with overall, cumulative, by_stratum, parameters.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_detection_probability(d, 0.01)
```

surv_estimate

Pipe-friendly surveillance analysis

Description

Convenience wrapper that creates a design, estimates prevalence, and optionally applies delay correction in a single pipe-friendly call. Designed for rapid exploratory analysis in interactive sessions.

Usage

```
surv_estimate(  
  data,  
  strata,  
  sequencing_rate,  
  population,  
  lineage,  
  correct_delay = FALSE,  
  method = "hajek",  
  ...  
)
```

Arguments

data	Data frame of sequence records.
strata	One-sided formula for stratification.
sequencing_rate	Sequencing rate specification (see surv_design()).
population	Population data frame.
lineage	Character. Target lineage to estimate.
correct_delay	Logical. Apply delay correction? Default FALSE.
method	Character. Prevalence method. Default "hajek".
...	Additional arguments passed to surv_design() .

Value

A `surv_prevalence` or `surv_adjusted` object.

See Also

[surv_design\(\)](#), [surv_lineage_prevalence\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
# One-liner analysis:
result <- surv_estimate(
  data = sim$sequences, strata = ~ region,
  sequencing_rate = sim$population[c("region", "seq_rate")],
  population = sim$population,
  lineage = "BA.2.86"
)
print(result)
```

surv_filter

Subset a surveillance design by filter criteria

Description

Creates a new `surv_design` object containing only sequences matching the specified filter criteria.

Usage

```
surv_filter(design, ...)
```

Arguments

design	A <code>surv_design</code> object.
...	Filter expressions passed to dplyr::filter() .

Value

A new surv_design object with filtered data.

See Also

[surv_design\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 5, n_weeks = 12, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
d_sub <- surv_filter(d, region %in% c("Region_A", "Region_B"))
print(d_sub)
```

surv_naive_prevalence *Compute naive (unweighted) lineage prevalence*

Description

Simple proportion without design correction. Useful as baseline.

Usage

```
surv_naive_prevalence(design, lineage, time = "epiweek", conf_level = 0.95)
```

Arguments

design	A surv_design object.
lineage	Character. Target lineage.
time	Character. Time aggregation. Default "epiweek".
conf_level	Numeric. Default 0.95.

Value

A surv_prevalence object with method = "naive".

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
naive <- surv_naive_prevalence(d, "BA.2.86")
```

surv_plot_allocation *Plot allocation plan*

Description

Plot allocation plan

Usage

```
surv_plot_allocation(allocation)
```

Arguments

allocation A surv_allocation object.

Value

A ggplot2 object.

Examples

```
sim <- surv_simulate(n_regions = 5, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
a <- surv_optimize_allocation(d, "min_mse", total_capacity = 500)
surv_plot_allocation(a)
```

surv_plot_sequencing_rates

Plot sequencing rate inequality across strata

Description

Plot sequencing rate inequality across strata

Usage

```
surv_plot_sequencing_rates(design)
```

Arguments

design A surv_design object.

Value

A ggplot2 object.

Examples

```
sim <- surv_simulate(n_regions = 5, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_plot_sequencing_rates(d)
```

surv_power_curve	<i>Compute power curve for detection across prevalence range</i>
------------------	--

Description

Generates a detection probability curve that can be directly plotted or included in publications. Answers: "At what prevalence does our surveillance achieve X% detection?"

Usage

```
surv_power_curve(
  design,
  prevalence_range = seq(0.001, 0.05, by = 0.001),
  delay_fit = NULL,
  thresholds = c(0.5, 0.8, 0.95)
)

## S3 method for class 'surv_power_curve'
plot(x, ...)
```

Arguments

design	A surv_design object.
prevalence_range	Numeric vector of prevalences to evaluate. Default seq(0.001, 0.05, by = 0.001).
delay_fit	Optional surv_delay_fit.
thresholds	Numeric vector of detection thresholds to mark. Default c(0.5, 0.8, 0.95).
x	A surv_power_curve object.
...	Additional arguments (unused).

Value

A list with:

curve Tibble with prevalence and detection columns.

thresholds Tibble with threshold, prevalence_needed columns.

A ggplot2 object.

See Also

[surv_detection_probability\(\)](#), [surv_required_sequences\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
pc <- surv_power_curve(d)
pc$thresholds
```

surv_prevalence_by *Estimate prevalence by subgroup*

Description

Applies [surv_lineage_prevalence\(\)](#) within subgroups defined by a grouping variable. Analogous to [survey::svyby\(\)](#) for stratified survey analysis.

Usage

```
surv_prevalence_by(
  design,
  lineage,
  by,
  time = "epiweek",
  method = "hajek",
  conf_level = 0.95
)
```

Arguments

design	A <code>surv_design</code> object.
lineage	Character. Target lineage.
by	Character. Column name to group by (e.g., "region" or "source_type").
time	Character. Time aggregation. Default "epiweek".
method	Character. Estimation method. Default "hajek".
conf_level	Numeric. Default 0.95.

Value

A tibble with columns: group, time, lineage, prevalence, se, ci_lower, ci_upper, n_obs, effective_n.

See Also

[surv_lineage_prevalence\(\)](#), [surv_filter\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 4, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")],
                 sim$population, source_type = "source_type")
surv_prevalence_by(d, "BA.2.86", by = "region")
```

surv_quality

Compute surveillance quality metrics

Description

Returns a single-row tibble of design quality indicators suitable for inclusion in manuscripts. Analogous to `broom::glance()` but for the surveillance design itself.

Usage

```
surv_quality(design, target_lineage = NULL, target_prevalence = 0.01)
```

Arguments

`design` A `surv_design` object.

`target_lineage` Character or NULL. Default NULL auto-selects.

`target_prevalence` Numeric. For detection calculation. Default 0.01.

Value

A single-row tibble with columns: `n_obs`, `n_strata`, `gini`, `rate_ratio`, `effective_n`, `deff`, `detection_prob`, `mean_bias`.

See Also

[surv_report\(\)](#), [surv_design\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_quality(d)
```

surv_report	<i>Generate a comprehensive surveillance system report</i>
-------------	--

Description

Produces a summary of the current surveillance design's strengths, weaknesses, and recommendations.

Usage

```
surv_report(design, target_lineage = NULL, target_prevalence = 0.01)
```

Arguments

`design` A `surv_design` object.

`target_lineage` Character or NULL. Lineage to focus on. If NULL, uses the most common non-"Other" lineage.

`target_prevalence` Numeric. Assumed prevalence for detection calculations. Default 0.01.

Value

Invisibly returns a named list of computed metrics including `n_obs`, `n_strata`, `rate_range`, `gini`, `effective_n`, `detection_prob`, and `mean_bias`.

See Also

[surv_design\(\)](#), [surv_lineage_prevalence\(\)](#), [surv_detection_probability\(\)](#), [surv_optimize_allocation\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_report(d)
```

surv_reporting_probability

Compute cumulative reporting probability

Description

Compute cumulative reporting probability

Usage

```
surv_reporting_probability(delay_fit, delta, stratum = NULL)
```

Arguments

delay_fit	A surv_delay_fit object.
delta	Integer vector. Days since collection.
stratum	Character or NULL. Default NULL.

Value

Numeric vector of probabilities.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 12, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                sim$population[c("region", "seq_rate")], sim$population)
fit <- surv_estimate_delay(d)
surv_reporting_probability(fit, delta = c(7, 14, 21))
```

surv_required_sequences

Required sequences for target detection probability

Description

Required sequences for target detection probability

Usage

```
surv_required_sequences(
  true_prevalence,
  target_probability = 0.95,
  n_periods = 1L,
  detection_threshold = 1L
)
```

Arguments

true_prevalence Numeric.
 target_probability Numeric. Default 0.95.
 n_periods Integer. Default 1.
 detection_threshold Integer. Default 1.

Value

Integer.

Examples

```

surv_required_sequences(0.01)
surv_required_sequences(0.05, target_probability = 0.99)

```

surv_sensitivity *Sensitivity analysis across methods*

Description

Runs all three prevalence estimators and delay/no-delay variants on the same design, producing a comparison table. Essential for robustness checks in publications.

Usage

```

surv_sensitivity(
  design,
  lineage,
  delay_fit = NULL,
  time = "epiweek",
  conf_level = 0.95
)

```

Arguments

design A surv_design object.
 lineage Character. Target lineage.
 delay_fit Optional surv_delay_fit object. If provided, includes delay-adjusted estimates.
 time Character. Default "epiweek".
 conf_level Numeric. Default 0.95.

Value

A tibble with one row per method-time combination, columns: method, time, prevalence, se, ci_lower, ci_upper.

See Also

[surv_lineage_prevalence\(\)](#), [surv_adjusted_prevalence\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
surv_sensitivity(d, "BA.2.86")
```

<code>surv_set_weights</code>	<i>Override design weights with custom values</i>
-------------------------------	---

Description

Override design weights with custom values

Usage

```
surv_set_weights(design, weights)
```

Arguments

<code>design</code>	A <code>surv_design</code> object.
<code>weights</code>	Numeric vector of length equal to number of strata.

Value

Updated `surv_design`.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 4, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
d2 <- surv_set_weights(d, rep(1.0, d$n_strata))
```

 surv_simulate

Simulate genomic surveillance data

Description

Generates synthetic surveillance datasets with realistic features: multiple regions with unequal sequencing rates, multiple lineages with time-varying prevalence, configurable reporting delays, and multiple sample sources.

Usage

```
surv_simulate(
  n_regions = 5L,
  n_weeks = 26L,
  total_positive_per_week = 1000L,
  sequencing_rates = NULL,
  lineage_dynamics = NULL,
  delay_params = list(mu = 10, size = 3),
  sources = c("clinical", "wastewater", "sentinel"),
  source_weights = c(0.7, 0.2, 0.1),
  seed = NULL
)
```

Arguments

n_regions	Integer. Number of geographic regions. Default 5.
n_weeks	Integer. Number of epiweeks. Default 26.
total_positive_per_week	Integer. Mean total positive cases per week across all regions. Default 1000.
sequencing_rates	Numeric vector of length n_regions. Per-region sequencing probability. If NULL, generated from a Beta distribution with realistic inequality. Default NULL.
lineage_dynamics	Named list of functions, each taking a week number and returning a positive weight. If NULL, uses a default four-lineage scenario. Default NULL.
delay_params	List with mu and size for negative binomial reporting delay. Default list(mu = 10, size = 3).
sources	Character vector of sample source types. Default c("clinical", "wastewater", "sentinel").
source_weights	Numeric vector (same length as sources). Default c(0.7, 0.2, 0.1).
seed	Integer or NULL. Random seed. Default NULL.

Value

A named list with elements:

sequences Tibble of individual sequence records.

population Tibble with one row per region.

truth Tibble of true lineage prevalence by region and week.

parameters List of all input parameters.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 8, seed = 42)
head(sim$sequences)
sim$population
```

surv_table

Format prevalence results for knitr tables

Description

Produces a publication-ready summary table from any survinger result. Automatically called when objects are printed inside RMarkdown chunks if knitr is loaded.

Usage

```
surv_table(x, digits = 3, percent = TRUE, ...)
```

Arguments

x	A survinger result object.
digits	Integer. Decimal places for prevalence. Default 3.
percent	Logical. Display as percentages? Default TRUE.
...	Additional arguments (unused).

Value

A tibble formatted for display.

See Also

[surv_lineage_prevalence\(\)](#), [tidy.surv_prevalence\(\)](#)

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
prev <- surv_lineage_prevalence(d, "BA.2.86")
surv_table(prev)
```

surv_update_rates *Update sequencing rates in a surveillance design*

Description

Update sequencing rates in a surveillance design

Usage

```
surv_update_rates(design, new_rates)
```

Arguments

design A surv_design object.

new_rates Data frame with strata columns + seq_rate, or named numeric vector.

Value

Updated surv_design.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 4, seed = 1)
d <- surv_design(sim$sequences, ~ region,
                 sim$population[c("region", "seq_rate")], sim$population)
new_r <- sim$population[c("region", "seq_rate")]
new_r$seq_rate <- new_r$seq_rate * 2
d2 <- surv_update_rates(d, new_r)
```

theme_survinger *Publication-quality ggplot2 theme*

Description

A clean, high-contrast theme suitable for journal publication. Follows Nature/Lancet figure guidelines.

Usage

```
theme_survinger(base_size = 11, base_family = "")
```

Arguments

`base_size` Base font size. Default 11.
`base_family` Font family. Default "".

Value

A ggplot2 theme object.

See Also

[surv_compare_estimates\(\)](#)

Examples

```
library(ggplot2)  
ggplot(mtcars, aes(wt, mpg)) + geom_point() + theme_survinger()
```

tidy.surv *Extract tidy estimates from survinger objects*

Description

Converts survinger result objects into tidy tibbles suitable for further analysis with dplyr, ggplot2, or other tidyverse tools.

Usage

```
## S3 method for class 'surv_prevalence'  
tidy(x, ...)  
  
## S3 method for class 'surv_nowcast'  
tidy(x, ...)  
  
## S3 method for class 'surv_adjusted'  
tidy(x, ...)  
  
## S3 method for class 'surv_allocation'  
tidy(x, ...)  
  
## S3 method for class 'surv_delay_fit'  
tidy(x, ...)
```

Arguments

x	A survinger result object.
...	Additional arguments (currently unused).

Value

A tibble.

Examples

```
sim <- surv_simulate(n_regions = 3, n_weeks = 10, seed = 1)  
d <- surv_design(sim$sequences, ~ region,  
                 sim$population[c("region", "seq_rate")], sim$population)  
prev <- surv_lineage_prevalence(d, "BA.2.86")  
tidy(prev)
```

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